

Provenance of recycled palynomorph assemblages recovered from surficial glaciomarine sediments in Bransfield Strait, offshore Antarctic Peninsula

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Abstract

Glaciomarine surficial sediments in cores taken from Bransfield Strait, adjacent to the Antarctic Peninsula, have yielded abundant recycled and contemporaneous (Pleistocene–Holocene) palynomorphs. The former are derived principally from Late Cretaceous–Palaeogene sediments and provide information on glaciomarine depositional conditions and sediment source areas. The composition of the assemblages suggests that they reflect vegetation that was endemic to the Weddellian Biogeographic Province, which includes what is now Seymour Island, James Ross Island and other parts of the Antarctic Peninsula region. The sediments concerned are considered to have accumulated as a result of ice-rafting and discharge of subglacial meltwater.

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1. Introduction

The glaciomarine sediments that cover the sea-floor around the Antarctic Peninsula are rich in Recent and recycled marine and non-marine organic- and inorganic-walled microfossils, e.g., spores, pollen grains and dinoflagellate cysts, and diatoms and silicoflagellates respectively. These microfossils derive from sedimentary rocks that crop out either on the peninsula and neighbouring islands, where they are commonly hidden by thick ice, or at submarine locations on the continental shelf.

They accumulated as a result of glacial and/or submarine processes.

The occurrence of fossil palynofloras (organic-walled microfossils) in essentially modern glacial marine muds close to Antarctica has been reported previously on a number of occasions (e.g., Truswell, 1983; Truswell and Drewry, 1984; Byun et al., 1996). The specimens recovered have been used for determining the ages of rocks hidden beneath the ice around Antarctica (Kemp, 1972; Truswell, 1982, 1983, 1990; Truswell and Drewry, 1984) and suggesting the general composition of the vegetation that once clothed the continent. Cretaceous and Palaeogene palynomorphs are known from the eastern side of the Antarctic Peninsula, where outcrop successions on James Ross and Vega islands (Askin, 1983), Seymour Island (Askin, 1988a,b, 1989, 1990a,b, 1997), and

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in the James Ross Island Basin (Dettmann and Thomson, 1987) have been examined. Some of the Antarctic Cretaceous and Cenozoic spore and pollen (miospore) assemblages contain species that may have been produced by plant taxa that were endemic to the region, and some of the species are thought to have their earliest occurrences in the Antarctica, both implying that the parent plants originated in the region (Askin, 1989). In this paper we concentrate on the occurrence of recycled miospores and dinoflagellate cysts in the sea-floor sediments of the Bransfield Strait and their likely provenance.

2. Physiography and vegetation of the Antarctic Peninsula

The Bransfield Strait is a semi-enclosed sea about 300 km long and 100 km wide located between the South Shetland archipelago and the north-eastern tip of the Antarctic Peninsula (i.e., ca. 61–64°S; Fig. 1). This strait is a narrow, deep (up to >2000 m), volcanic, rifted basin (the Bransfield Basin) that has been actively spreading during the past 4 myr in response to subduction in the South Shetland Trench (Barker, 1982; González-Ferrán, 1985; Gamboa and Maldonado, 1990). The north-eastern part of the strait opens out into the Scotia and Weddell seas. The south-western part is connected to the eastern Bellingshausen Sea via the Gerlache Strait.

The north-east–south-west-orientated Bransfield Basin is divided into three subbasins: Western, Central and Eastern. These are separated from each other by sills at depths in the region of 500 m and progressively deepen north-eastward (Fig. 1A). The Western Subbasin, in which Core S7 was drilled, is irregular in shape, and flanked by shelves 20–50 km wide. The Central Subbasin, from which Cores S4 and AB2 were taken (the latter on its western margin), is more regular in shape with a relatively wide continental margin (ca. 100 km) on the Antarctic Peninsula side and, in contrast, a narrow shelf (<20 km) and steep slope (5–10°) on the margin of the South Shetland Islands (Prieto et al., 1997; Yoon, S. et al., 2002). The depth of the sea-floor in this subbasin increases gradually towards the north-east up to a little over 2000 m. The width of southern continental shelf is wider than that of the north owing to the north–south asymmetry of the Bransfield Basin. The Eastern Subbasin, which also extends north-eastward, has relatively wide continental margins (50–125 km), and reaches a depth of 2500 m.

Surface water circulation in the Bransfield Basin is controlled by the interaction of two different inflows: cold, saline Weddell Sea water on the Antarctic Peninsula side, and relatively warmer, less saline Bellingshausen Sea water on the South Shetland Islands side (López et al., 1999). Circulation at depth is mainly linked to the entrance of dense Weddell shelf waters around the north-east tip of Joinville Island, which then sinks over the shelf and down the slope into the basin (Whitworth et al., 1994; López et al., 1999).

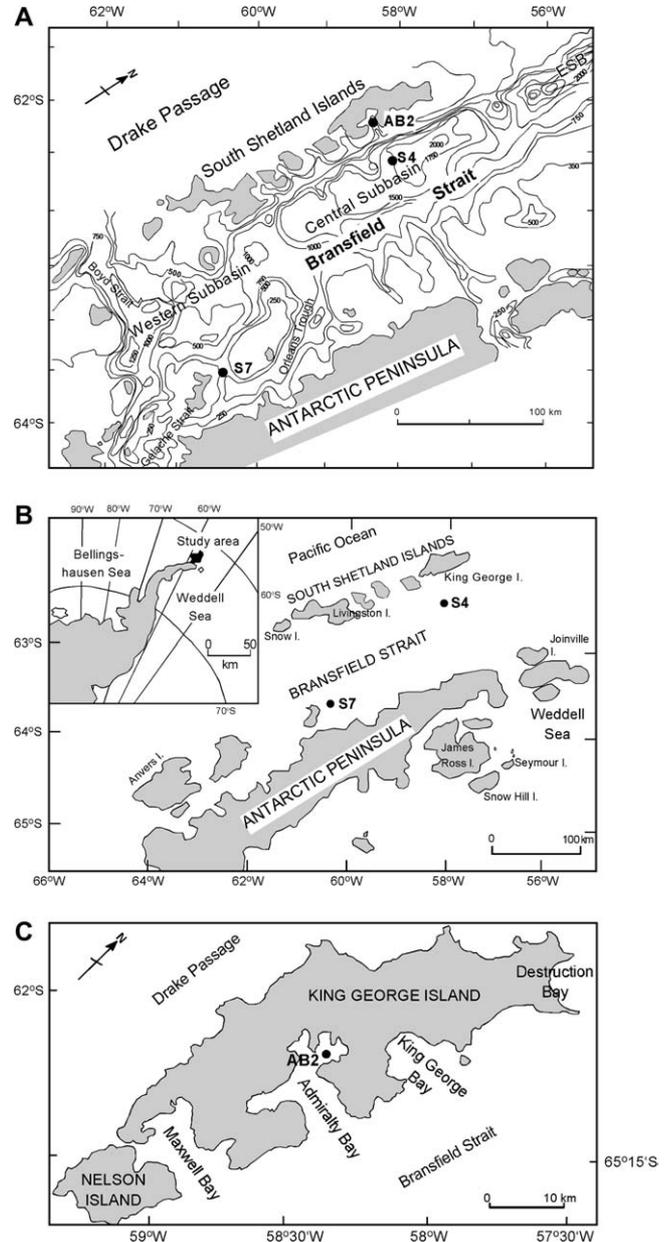


Fig. 1. A, bathymetric map of Bransfield Strait (from Jeffers and Anderson, 1991) with the location of cores; contours in metres; ESB, Eastern Subbasin. B, map showing the location of the study area (inset) and islands mentioned in the text. C, outline map of King George Island and neighbouring Nelson Island with the location of Core AB2 indicated.

The surficial sediments of the basin are composed mainly of sandy silt (4–6 Φ) and silty clay (>6 Φ). The latter is distributed over the deepest parts of the basin, whereas sandy silt deposits occur on the continental shelf and slope (Yoon, 1995).

Today vegetation on the Antarctic Peninsula is sparse, dominated by non-vascular plants, and mostly near the coast (Bliss, 1979). At least 200 species of lichens and 85 species of mosses together with many species of algae have been recorded from Antarctica (Walton and Bonner, 1985). Commonly colonizing rock surfaces, with some herbs on the

soils, all have to tolerate the harsh tundra environment. Representatives of only two flowering plant families, a pearlwort and a grass species, have been reported (Bliss, 1979; Longton, 1985), both of which are confined to the maritime zone.

3. Material and methods

During the 1991/92 survey season, the Korea Ocean Research and Development Institute (KORDI) operated a “Korean Antarctic Research Program” and drilled several piston cores in the Bransfield Basin, three of which were intended for palynological analysis. Sixty-eight samples were taken from these cores at intervals of 10–20 cm: 10 from Core S7, 31 from Core S4, and 27 from Core AB2.

Core S7 (Latitude 63°30'05"S, Longitude 60°26'06"W; Fig. 2) was drilled in water 675 m deep on the southern slope of the Western Subbasin. It is 523 cm long and consists of clay, silt, sand and gravel. Samples were not taken from the lower part of the section because of the high sand and gravel content, which is unsuitable for the preservation of palynomorphs. Core S4 (62°30'58"S, 58°05'58"W; Fig. 3), drilled in water 1846 m deep in the central part of the Central Subbasin, is 537 cm long and composed mainly of clay and silt, with a small amount of sand. Core AB2 (62°05'09"S, 58°23'06"W; Fig. 4), drilled in water 261 m deep in Admiralty Bay, King George Island, is 256 cm long and consists largely of mud (Park et al., 1992).

About 30 g of each of the dried samples were processed for palynological analysis using standard procedures (Moore et al., 1991) involving HCl (30%) and HF (45%) for the removal of calcareous and siliceous minerals. The organic matter recovered was oxidized with NaClO₂ (3%) to remove fine humic matter, and then sieved through screens of 10 µm mesh. The sieved residue was strewn mounted on glass slides in glycerine jelly. Raw data were converted into percentages using TILIA v2.0 (Grimm, 1993). TILIAGRAPH v1.25 (Grimm, 1991) was used to generate the ‘pollen’ diagrams. The percentages of recycled and contemporaneous spores and pollen grains were calculated from the total palynomorph sum (Figs. 2–4).

4. Results

All of the palynological assemblages examined contain Pleistocene–Holocene and recycled miospores. Selected examples of both are shown in Figs. 5–7. The younger taxa consist of cryptogam spores (*Cyathea* and Polypodiaceae) and pollen grains of herbs (Chenopodiaceae, Compositae, Ericaceae and Poaceae; Fig. 5). Among the recycled miospores are: the spores *Baculatisporites comaumensis*, *Ceratosporites equalis*, *Clavifera triplex*, *Cyatheacidites annulatus*, *Osmundacidites wellmanii* and *Retitriletes eminulus*; the gymnosperm pollen *Dacrycarpites australiensis*, *Lygistepollenites florinii*, *Microcachrydites antarcticus*, *Phyllocladites mawsonii*, *Phyllocladites palaeogenicus*, *Piceapollenites* sp., *Pinuspollenites* sp., *Podocarpidites marwickii*, *Beaupreacidites elegansiformis*, *Heliporites astrus*, *Nothofagidites emarcitius*, *Nothofagidites endurus*, *Nothofagidites incrassatus*, *Retistephanocolpites crassimurus*, *Tricopites reticulatus*, *Tripoporollenites chnosus*, *Tubifloridites antipodica*, Chenopodiaceae, *Cyathea*, Ericaceae, Poaceae, Recycled spores, Recycled gymnosperm pollen, Recycled angiosperm pollen, Younger pollen, Diversity, Abundance

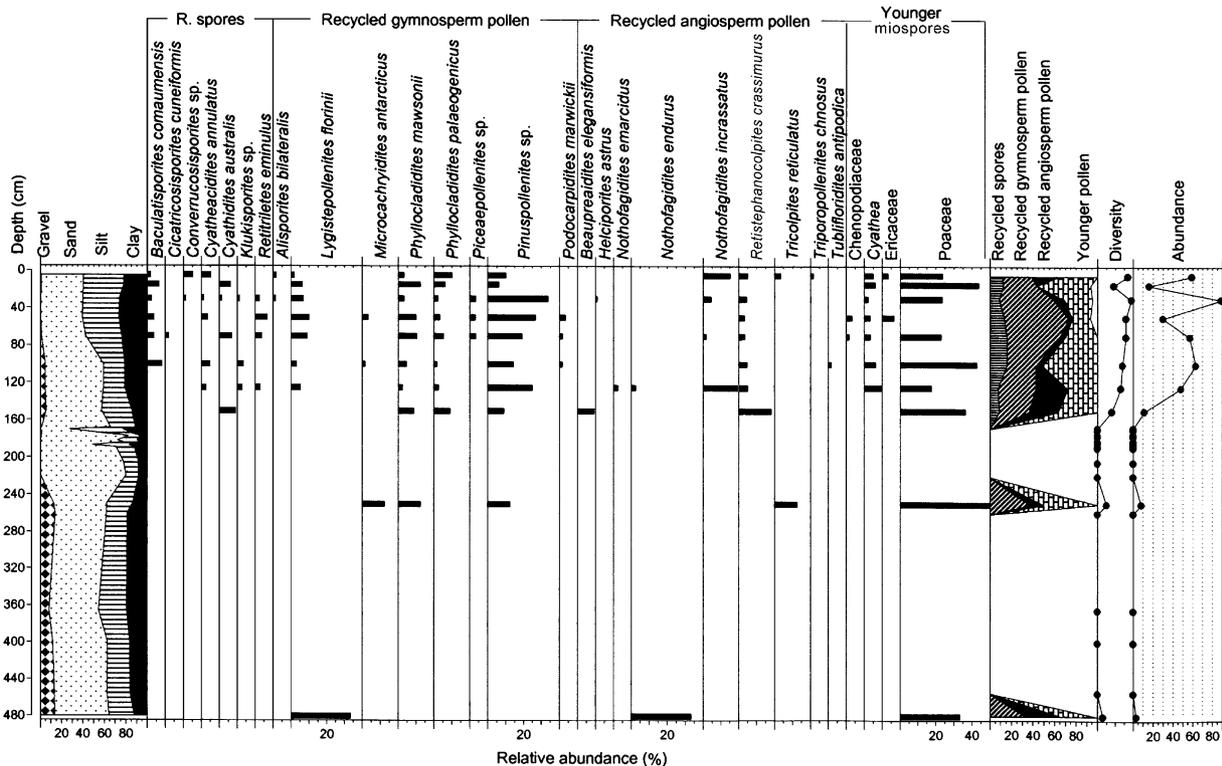


Fig. 2. ‘Pollen diagram’ of S7 core from Bransfield Strait showing most of the taxa recorded. Grain texture analysis is from Park et al. (1992).

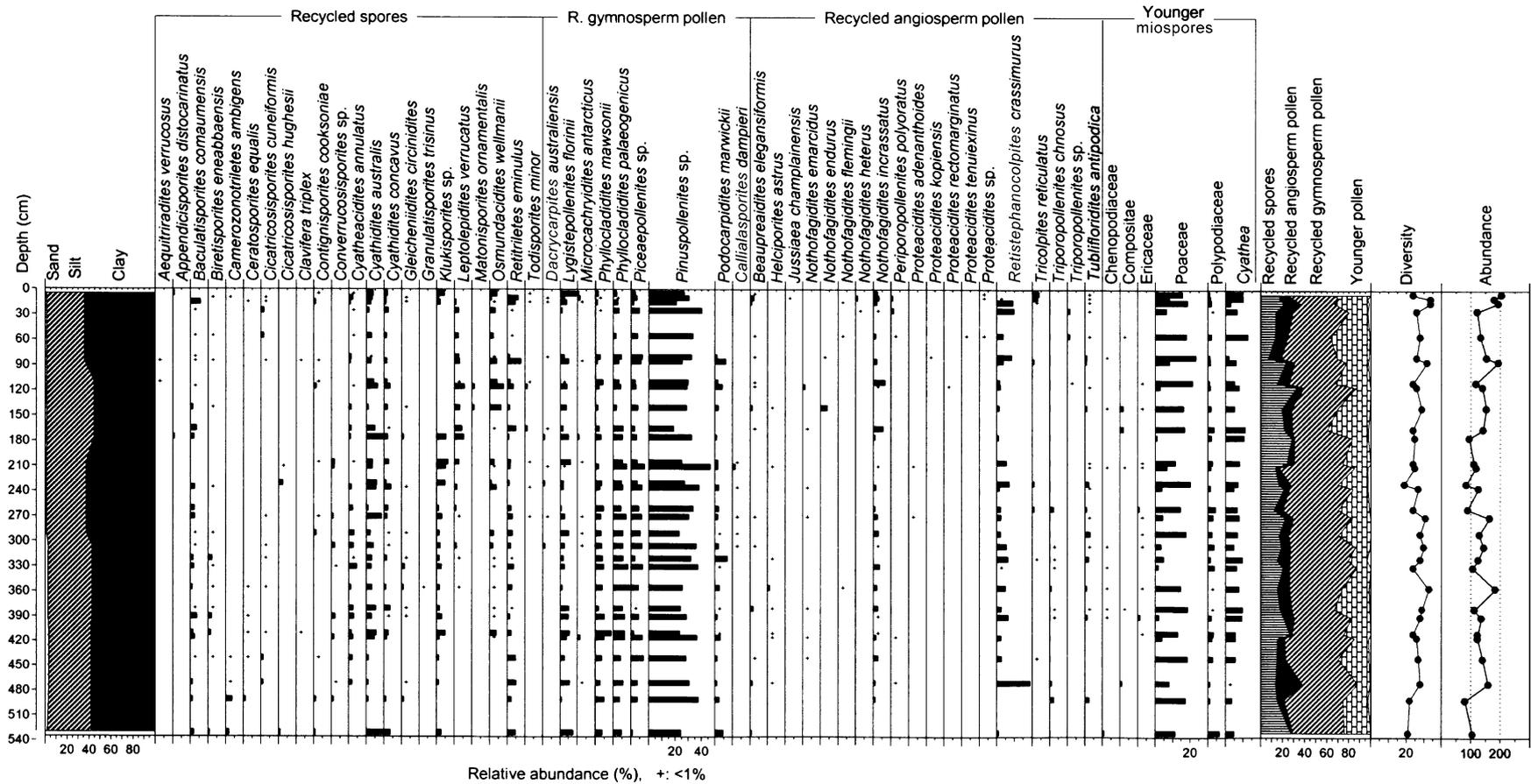


Fig. 3. 'Pollen diagram' of S4 core from Bransfield Strait showing most of the taxa recorded. Grain texture analysis is from the Park et al. (1992).

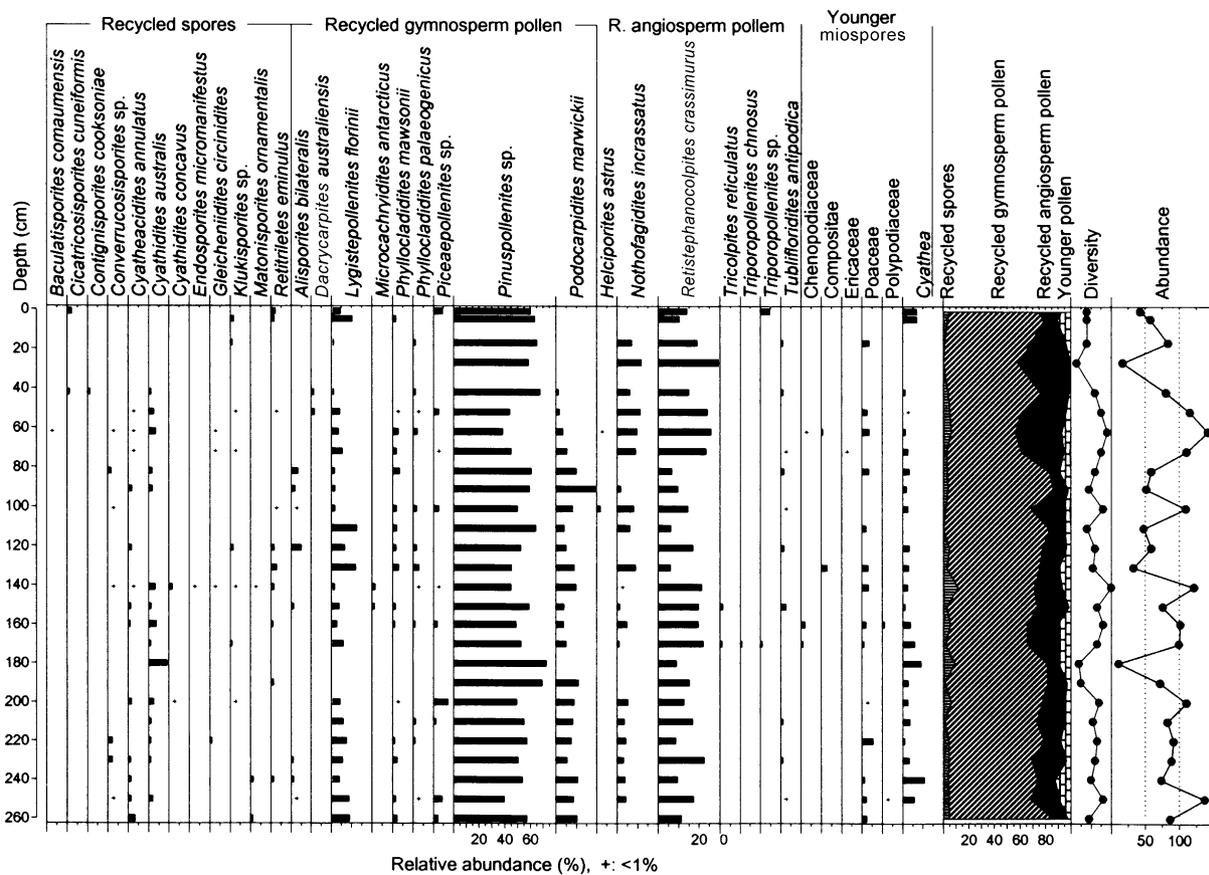


Fig. 4. 'Pollen diagram' of AB2 core from Bransfield Strait showing most of the taxa recorded. Grain texture analysis is from Park et al. (1992).

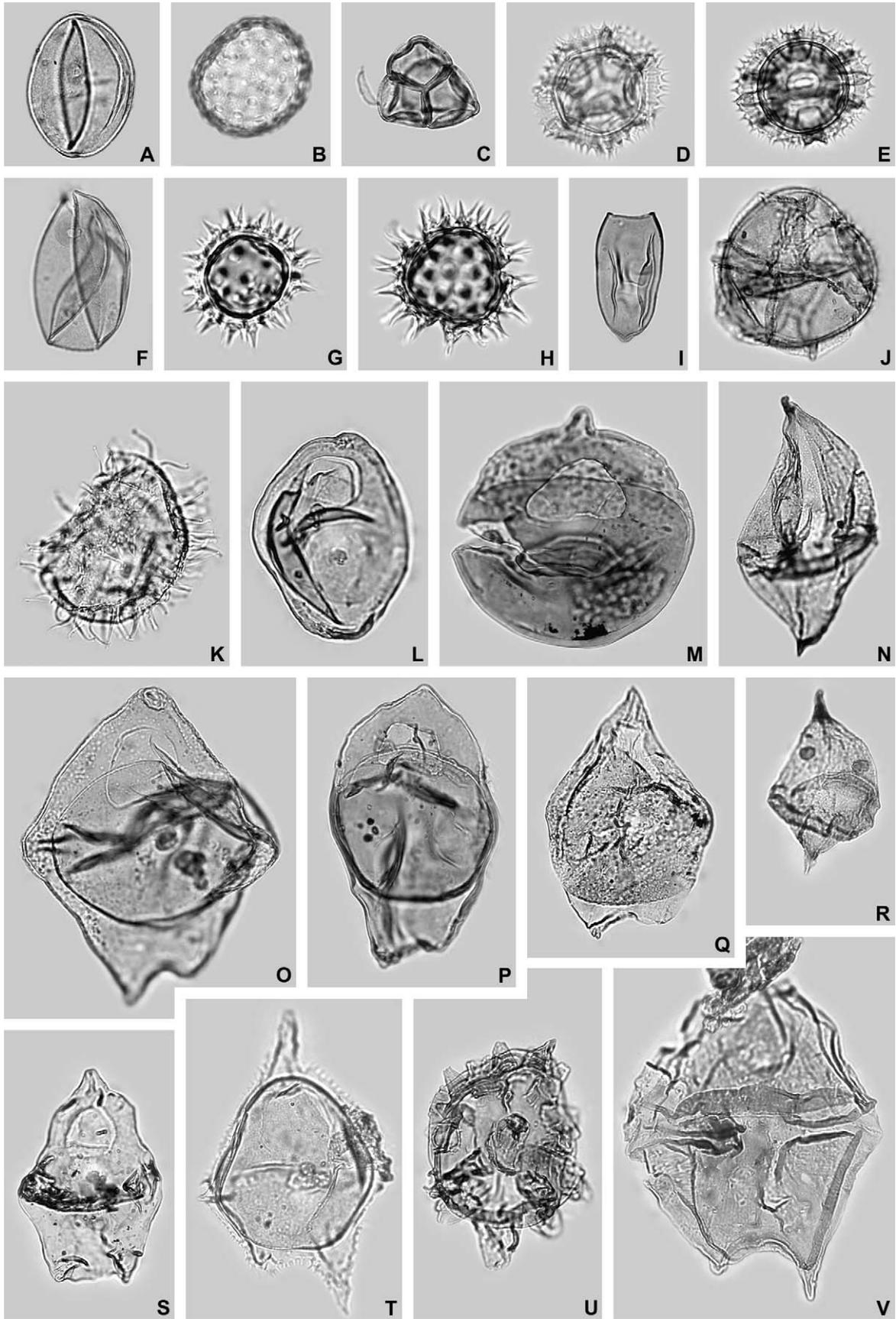
angiosperm pollen *Beaupreaidites elegansiformis*, *Nothofagidites endurus*, *N. flemingii*, *N. incrassatus*, *Periporopollenites polyoratus*, *Proteacidites adenanthoides*, *P. kopiensis*, *P. rectomarginatus*, *P. tenuixinus*, *Tricolpites reticulatus*, *Tripoporopollenites chnosus* and *Tubulifloridites antipodica* (Figs. 6, 7, Table 1).

In Core S7, recycled gymnosperm pollen grains *Lygistepollenites florinii*, *Phyllocladidites mawsonii*, *P. palaeogenicus* and *Pinuspollenites* sp. are especially common along with reworked angiosperm pollen referable to *Retistephanocolpites crassimurus*. Contemporaneous products include abundant (ca. >20%) representatives of the Poaceae and common spores attributable to *Cyathea*. Other common, or intermittently numerous, taxa include recycled *Baculatisporites comauensis*, *Cyathidites australis* and *Nothofagidites incrassatus* (Fig. 2).

In Core S4, the palynological assemblages are dominated (ca. 5–40%) by recycled gymnosperm pollen (*Pinuspollenites* sp.) and younger angiosperm pollen (Poaceae), in combination with common (ca. <5%) recycled *Cyathidites australis*, *Osmundacidites wellmanii* and *Retitriletes eminulus* and contemporaneous spores of *Cyathea*. Also common are recycled *Lygistepollenites florinii*, *Nothofagidites incrassatus*, *Phyllocladidites mawsonii*, *P. palaeogenicus*, *Piceapollenites* sp. and *Retistephanocolpites crassimurus* (Fig. 3).

In Core AB2, recycled *Pinuspollenites* sp. and *Retistephanocolpites crassimurus* dominate (ca. 20–60%) throughout the section. Among the common taxa, reworked *Lygistepollenites florinii*, *Nothofagidites incrassatus* and *Podocarpidites marwickii* are especially abundant along with comparatively numerous spores of *Cyathea* and representatives of the Poaceae (Fig. 4).

Fig. 5. Examples of contemporaneous (Pleistocene–Holocene) angiosperm pollen grains (A–H), acritarch (I), and dinoflagellate cysts (J, K), and recycled dinoflagellate cysts (L–V) in the core samples from Bransfield Strait examined. The specimens are identified by core, sample depth (cm) and England Finder reference. A, Poaceae, Core S4, 55–60, F27/3 + 4. B, Chenopodiaceae, Core S4, 210–220, F17/3 + 4. C, Ericaceae, Core S4, 270–280, E20/3. D, E, Compositae: D, Core S7, 30–35, B42/2; E, Core S4, 320–325, F44/4. F, Poaceae, Core S4, 55–60, F27/3 + 4. G, H, Compositae: G, Core S4, 320–325, G17/2; H, Core S4, 55–60, N37/1 + 3. I, *Beringiella fritilla*, Core AB2, 52–57, E45/4. J, *Impagidinium patulum*, Core AB2, 141–146, V27. K, *Operculodinium centrocarpum*, Core S4, 140–145, U37. L, *Manumiella? cretacea*, Core S4, 55–60, L26. M, *Nelsoniella tuberculata*, Core S4, 290–295, Q24/1. N, *Diconodinium cristatum*, Core S4, 390–400, G28/1 + 3. O, *Manumiella seelandica* (Lange, 1969) Bujak and Davies, 1983, emend. Firth, 1987, Core S4, 115–125, C38/1. P, *Isabelidinium* sp., Core S4, 330–340, T19/3 + 4. Q, *Isabelidinium pellucidum*, Core S4, 85–95, C30/3 + 4. R, *Laciniadinium arcticum*, Core S4, 320–325, L20/1 + 3. S, *Chatangiella* sp., Core S4, 205–210, T20/3. T, *Senegalinium* sp., Core S4, 115–125, Y14/2. U, *Cassidinium fragile*, Core S4, 305–315, W25/1. V, *Paleoperidinium pyrophorum*, Core S4, 85–95, D14/1 + 2. A–H, $\times 900$; I–V, $\times 400$.



5. Discussion

5.1. Age of recycled and contemporaneous palynofloras

The ages represented by the contemporaneous terrestrial palynofloras cannot be determined precisely because most of the components have biostratigraphic ranges that extend from the Oligocene to the present day (e.g., Cookson, 1947; Germeraad et al., 1968). However, it is possible to draw some conclusions by taking account of their associations with microplankton that have been recorded from the same cores as well as with the results of studies on these and other cores taken for different purposes (micropalaeontology: Yi et al., 1995; Byun et al., 1996; Lee, 1996; Shin et al., 2001; sedimentology: Yoon, H. et al., 2000; Yoon, S. et al., 2002; Bahk et al., 2003; organic geochemistry: Khim et al., 2002).

The diatoms, silicoflagellates and dinoflagellate cysts indicate that the surficial sediments were deposited from the beginning of the Pleistocene onwards. The silicoflagellate assemblages recovered from Cores S7 and S4 compare well with those of the Pleistocene–Holocene *Distephanus speculum speculum* Zone (Yi et al., 1995). Lee (1996) identified two diatom zones, a *Thalassiosira lentiginosa* Partial Range Zone and an *Actinocyclus ingens* Partial Range Zone, in all three of the cores examined for their microfossil content, and concluded that they also indicate an age-range of Pleistocene–Holocene. This determination is further supported by the occurrence of the acritarch *Beringiella fritilla* and the dinoflagellate cyst species *Impagidinium pallidum* and *I. patulum* in the same cores (Byun et al., 1996) (Fig. 5). Other authors have also regarded the S7 and S4 successions as having accumulated during the Quaternary period (Yoon, H. et al., 2000; Khim et al., 2002; Yoon, S. et al., 2002; Bahk et al., 2003). As a result, all of the younger palynofloras recorded from the S7, S4 and AB2 cores can be dated as Pleistocene–Holocene.

Although there is evidence of a small Palaeozoic contribution, the age-range of the dominant components of the recycled palynofloral assemblages recovered can be ascertained by comparison with the composition of the Late Cretaceous–Palaeogene assemblages and zonations of Dettmann and Thomson (1987), Askin (1988a,b, 1990a,b), Dettmann and Jarzen (1988), Dettmann (1989) and others. A variety of cryptogam spores, araucarian pollen grains, the gymnospermous *Microcachrydites* and *Podocarpidites*, and

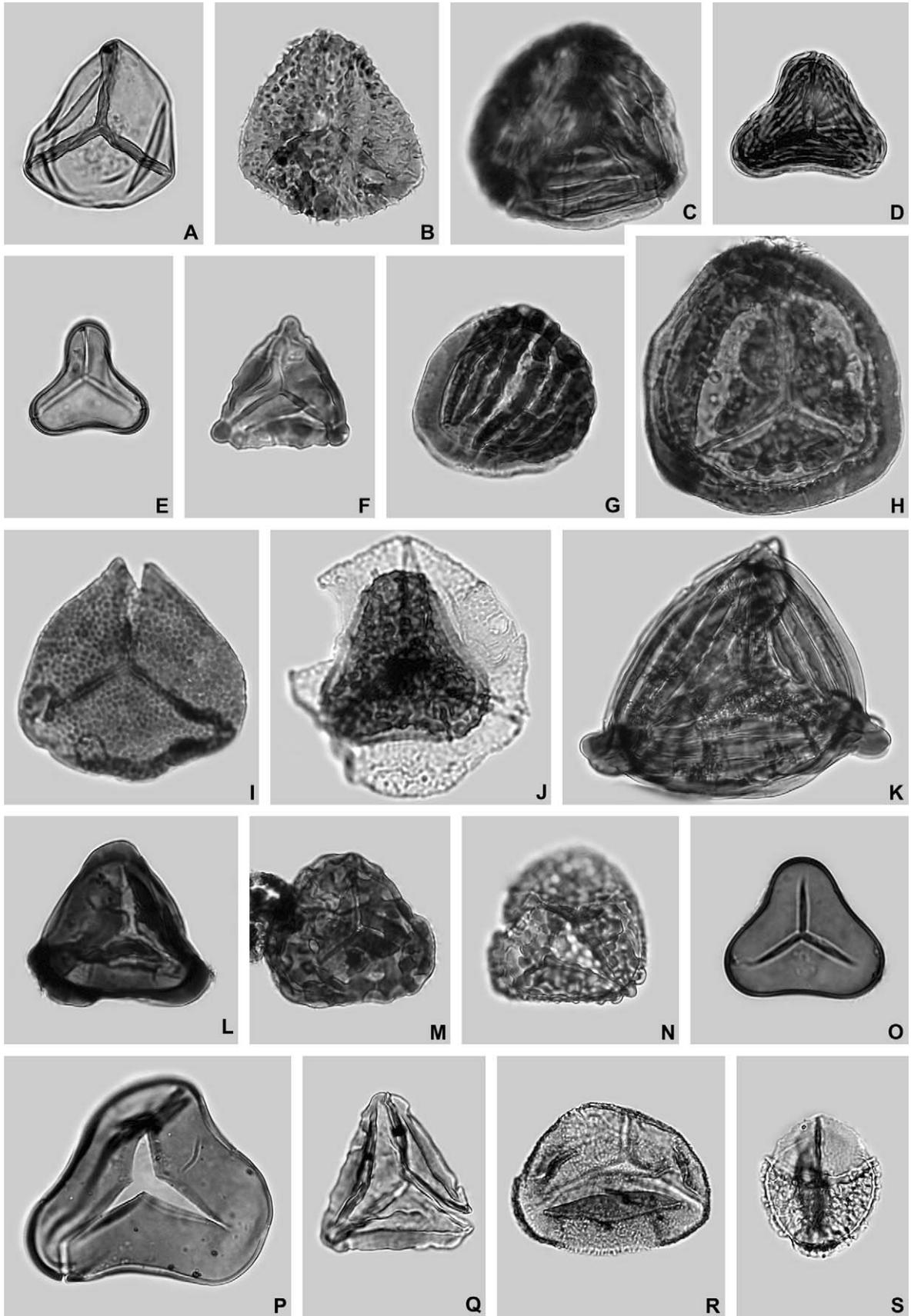
angiospermous *Beaupreaidites*, *Nothofagidites*, *Proteacidites*, *Tricolpites* and *Tubulifloridites* are common in Campanian–Maastrichtian sediments of the Antarctic Peninsula. Various proteaceous plant taxa were well-established in the Antarctic Peninsula area by the Late Campanian (Askin, 1983, 1989; Dettmann and Thomson, 1987; Dettmann and Jarzen, 1988; Dettmann, 1989). The Australasian–Antarctic region has been suggested as the place of origin of ‘*Beauprea*-type’ proteaceous plants during the Campanian (Dettmann and Jarzen, 1988; Pocknall and Crosbie, 1988). Angiosperm diversification reached a peak during the Campanian–Maastrichtian when a mixed conifer (podocarp-type and araucarian)-Proteaceae–*Nothofagus* rain forest clothed the Antarctic Peninsula (Askin, 1992). This vegetation formed part of the Late Cretaceous–Early Cenozoic Weddellian Biogeographic Province (Case, 1988; Fig. 8).

The spores and pollen grains recorded from the Bransfield Strait cores are common to the palynofloras of the Upper Campanian–Maastrichtian–Lower Danian Lopez de Bertodano Formation on Seymour Island (Askin, 1990a) and Maastrichtian deposits on Vega Island (Dettmann and Thomson, 1987). The recycled dinoflagellates that have been recorded from the cores (Fig. 5) have been dated as Late Cretaceous–early Tertiary (Palaeogene) by Byun et al. (1996). Accordingly, the reworked spores and pollen grains recorded in this study are considered to range in age between Late Campanian and early Palaeogene.

5.2. Potential provenance of recycled palynofloras

During the Cretaceous and Palaeogene the northern tip of the Antarctic Peninsula was situated at about 59–62°S (Lawver et al., 1985; Case, 1988) (Fig. 8). Throughout the Late Cretaceous, the peninsula area apparently underwent general uplift and was periodically part of an active volcanic arc adjacent to a subduction zone. Exposures of Late Cretaceous marine deposits are confined today to the north-eastern side of the peninsula (Elliot, 1985). During this period the exposed regions were forested, as indicated by the remains of in situ trees (Francis and Poole, 2002). To the east of the peninsula Cretaceous–Palaeogene sediments were deposited in the James Ross Basin, with sediment supply from the active magmatic arc to the west-north-west, now represented by the Antarctic Peninsula land mass (Pirrie et al., 1991, 1997). Plant debris from the vegetation that clothed the peninsula was washed into this basin and preserved in marine sediments (Francis, 1986).

Fig. 6. Examples of recycled spores, and one contemporaneous spore (O), in the core samples from Bransfield Strait examined. The specimens are identified by core, sample depth (cm) and England Finder reference. A, *Biretisporites eneabbaensis*, Core S4, 290–295, E43/3. B, *Ceratosporites* sp., Core S4, 355–360, D14. C, *Cicatricosisporites hughesii*, Core S4, 210–220, D20/1. D, *Cicatricosisporites cuneiformis*, Core S4, 55–60, X18/4. E, *Cyathidites concavus*, Core S4, 115–125, Q30/4. F, *Clavifera triplex*, Core S4, 410–415, N18/1 + 3. G, *Contignisporites cooksoniae*, Core S4, 5–10, X50/3 + 4 (distal view). H, *Cyatheacidites annulatus*, Core S4, 165–170, V40/3 + 4. I, *Granulatisporites trisinus*, Core S4, 355–360, V30/1. J, *Aequitriradites* sp. cf. *A. verrucosus*, Core S4, 530–537, O48/3. K, *Appendicisporites distocarينات*, Core S4, 175–185, X43/3. L, *Matonisporites agatonensis*, Core AB2, 240–245, E43/4. M, *Klukisporites* sp., Core AB2, 17–22, L41/1 + 3. N, *Leptolepidites verrucatus*, Core S4, 55–60, P19/3. O, *Cyathea*, Core S4, 230–235, Q26/2. P, *Cyathidites australis*, Core S4, 175–185, S38. Q, *Gleicheniidites circinidites*, Core S4, 140–145, Y27/3 + 4. R, *Osmundacidites* sp. cf. *O. wellmanii*, Core S4, 115–125, Q21/2 + 4. S, *Retitriletes eminus*, Core S4, 10–15, E14/4 (lateral view). All × 900.



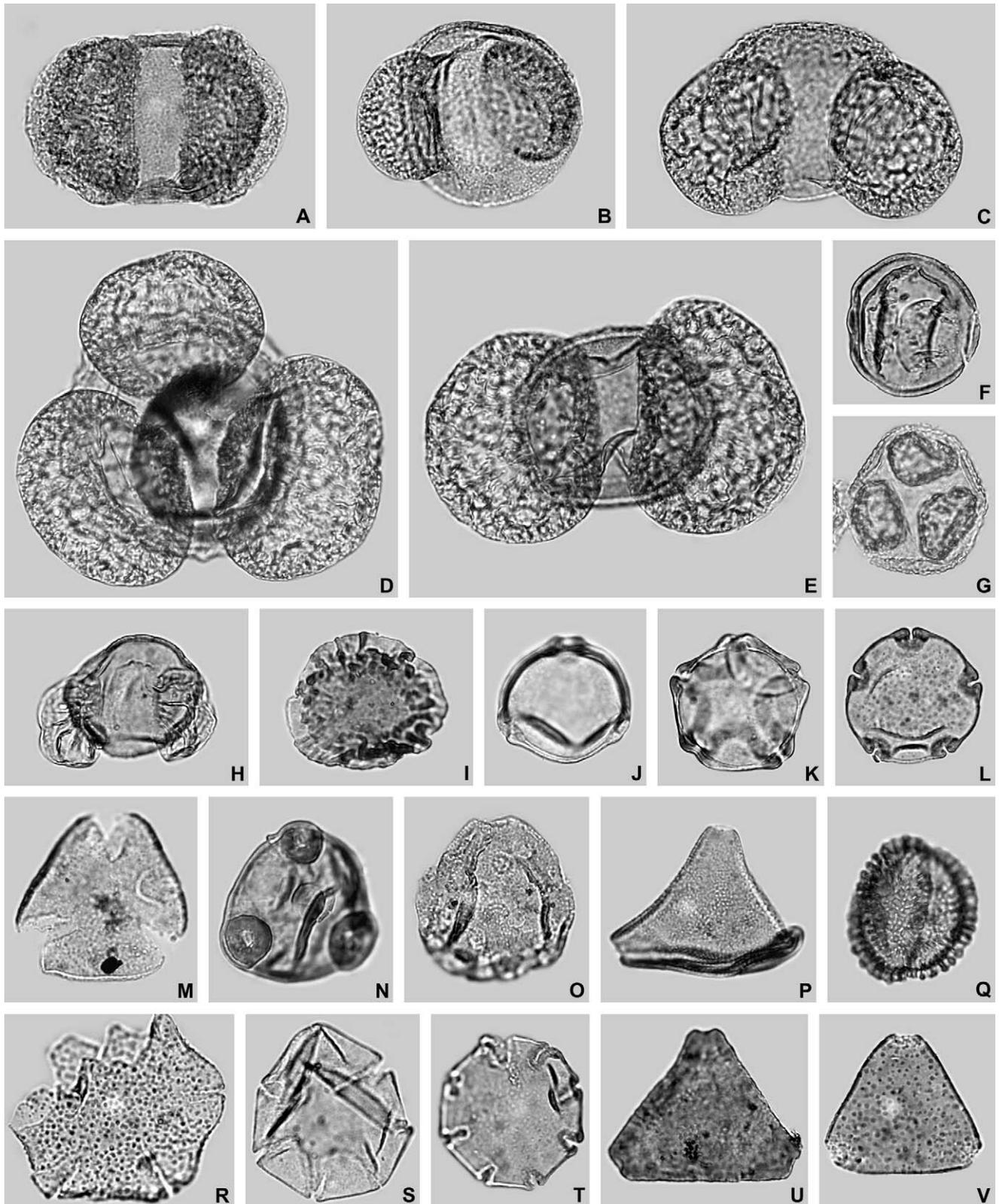


Fig. 7. Examples of recycled gymnosperm (A–I) and angiosperm (J–V) pollen grains in the core samples from Bransfield Strait examined. The specimens are identified by core, sample depth (cm) and England Finder reference. A, *Alisporites bilateralis*, Core AB2, 82–86, U13/4. B, *Lygistepollenites florinii*, Core S4, 175–185, Q21. C, *Pinuspollenites* sp., Core S4, 25–35, D11/4. D, *Dacrycarpites australiensis*, Core S4, 210–220, C24/2. E, *Podocarpidites marwickii*, Core S4, 305–315, O29/1 + 2. F, *Phyllocladites palaeogenicus*, Core S4, 110–115, H21/3. G, *Microcachrydites antarcticus*, Core S4, 5–10, D28/3. H, *Phyllocladites mawsonii*, Core S4, 410–415, X34. I, *Callialasporites dampieri*, Core S4, 205–210, G21/2. J, *Triplopollenites chnosus*, Core S4, 55–60, F30/4. K, *Helciporites astrus*, Core AB2, 101–106, B26. L, *Nothofagidites flemingii*, Core AB2, 17–22, X13. M, *Beaupreaidites elegansiformis*, Core S4, 115–125, Y36/3. N, *Jussiaea champlainensis*, Core S4, 10–15, Q44/3. O, *Periporopollenites polyoratus*, Core S4, 25–35, X26/1. P, *Proteacidites adenanthoides*, Core S4, 210–220, C21/4. Q, *Tricolpites reticulatus*, Core S4, 320–325, D30/3. R, *Nothofagidites emarcidus*, Core S4, 55–60, O41/4. S, *Nothofagidites heterus*, Core S4, 380–385, W24/4. T, *Nothofagidites incrassatus*, Core S4, 140–145, G36. U, *Proteacidites rectomarginatus*, Core S4, 115–125, K33/1. V, *Proteacidites tenuixinus*, Core S4, 55–60, W34/3. All $\times 900$.

Table 1

Recycled spores and pollen grains recorded from the core samples examined, with author attributions and year of publication provided for the species; stratigraphic ranges based on data from Antarctica, Australia, and southern Argentina as reported in particular by Askin (1983, 1989, 1990a,b), Dettmann and Thomson (1987), Dettmann and Jarzen (1988), Dettmann (1989) and Baldoni and Askin (1993) (bibliographic details are in the publications concerned and not included here)

Spores

Aequitriradites verrucosus (Cookson and Dettmann) Cookson and Dettmann, 1961: Berriasian–Albian

Appendicisporites distocarinaratus Dettmann and Playford, 1968: Cenomanian–Turonian

Baculatisporites comaumensis (Cookson) Potonié, 1956: Late Jurassic–Mid Oligocene

Biretisporites enebbaensis Backhouse, 1978: Late Tithonian–Late Aptian
Camerazonotriletes ambigens (Fradkina) Playford, 1971: Late Campanian–Late Maastrichtian

Ceratospores equalis Cookson and Dettmann, 1958: Late Jurassic–Danian

Cicatricosisporites cuneiformis Pocock, 1965: Late Albian–?Coniacian

Cicatricosisporites hughesii Dettmann, 1963: Aptian–?Cenomanian

Clavifera triplex (Bolchovitina) Bolchovitina, 1966: Campanian–Paleocene

Contignisporites cooksoniae (Balme) Dettmann, 1963: Jurassic

Converrucosisporites sp.

Cyatheacidites annulatus Cookson, 1947 ex Potonié, 1956: Neocomian–Miocene

Cyathidites australis Couper, 1953: Jurassic–Oligocene

Cyathidites concavus (Bolchovitina) Dettmann, 1963: Jurassic–Aptian

Endosporites micromanifestus Hacquebard, 1957: Late Fammenian–Visean

Gleicheniidites circinidites (Cookson) Dettmann, 1963: Late Jurassic–Eocene

Granulatisporites trisinus Balme and Hennelly, 1956: Permian

Klukisporites sp.

Leptolepidites verrucatus Couper, 1953: Late Jurassic–Albian

Matonisporites agatonensis Backhouse, 1978: Late Jurassic–Early Berriasian

Matonisporites ornamentalis (Cookson) Partridge, 1973: Eocene–Miocene

Osmundacidites wellmanii Couper, 1953: Jurassic–Early Oligocene

Retitriletes eminulus (Dettmann) Srivastava, 1977: Late Campanian–Early Oligocene

Todisporites minor Couper, 1958: Triassic–Palaeogene

Gymnosperm pollen grains

Alisporites bilateralis Rouse, 1959: Jurassic–Maastrichtian

Callialasporites dampieri (Balme, 1957) Sukh Dev, 1961: Neocomian–Albian

Dacrycarpites australiensis Cookson and Pike, 1953: Paleocene–Early Oligocene

Lygistipollenites florinii (Cookson and Pike) Stover and Evans, 1973: Santonian–Late Pliocene

Microcachryidites antarcticus Cookson, 1947: Campanian–Oligocene

Phyllocladites mawsonii Cookson, 1947: Campanian–Oligocene

Phyllocladites palaeogenicus (Cookson and Pike, 1954) Truswell, 1983: Paleocene–Pliocene

Piceapollenites sp.

Pinuspollenites sp.

Podocarpidites marwickii Couper, 1953: Campanian–Early Oligocene

Angiosperm pollen grains

Beaupreaidites elegansiformis Cookson, 1950: Maastrichtian–Miocene

Helciporites astrus Partridge, 1973: Eocene

Jussiaea champlainensis Traverse, 1955: Eocene

Nothofagidites emarcidus (Cookson) Harris, 1965: Paleocene–Early Miocene

Nothofagidites endurus Stover and Evans, 1973: Maastrichtian–Early Eocene

Nothofagidites flemingii (Couper) Potonié, 1960: Late Campanian–Oligocene

Nothofagidites heterus (Cookson) Stover and Evans, 1973: Late Oligocene–Early Miocene

Nothofagidites incrassatus (Cookson) Dettmann, 1990: Eocene

Periporipollenites polyoratus (Couper) Stover, 1973: Maastrichtian–Early Eocene

Proteacidites adenanthoides Cookson, 1950: Campanian–Early Oligocene

Proteacidites kopiensis Harris, 1972: Campanian–Eocene

Table 1 (continued)

Proteacidites rectomarginatus Cookson, 1950: Maastrichtian–Late Miocene

Proteacidites tenuixinus Stover, 1973: Maastrichtian–Early Oligocene

Proteacidites sp.

Retistephanocolpites crassimurus Rao and Ramanujam, 1982: Lower–Middle Miocene

Tricolpites reticulatus Cookson, 1947: Campanian–Paleocene

Triporipollenites chnosus Partridge, 1973: Late Eocene–Miocene

Triporipollenites sp.

Tubulifloridites antipodica Cookson, 1947: Campanian–Pliocene

Antarctic outcrops of Late Cretaceous and Palaeogene age are restricted to the peninsula region. Fossil plants and palynomorphs occur on various islands in the James Ross Basin; in the South Shetland Islands on Livingston, King George and adjacent small islands (Fig. 1B); and on Alexander, Adelaide, and Brabant islands. Terrigenous detritus from these regions is transported into the marine realm and onto the ocean floor in a number of ways, but principally by turbidity, surface and bottom currents, and ice.

Direct correlation of the resulting recycled assemblages to their source areas is difficult because most of the South Shetland Islands and Antarctic Peninsula are covered by glaciers, and igneous rocks are also widespread. Hence, stratigraphic knowledge is limited to comparatively small areas of sedimentary rocks that are free of ice cover. Most of the South Shetland Islands are composed of Late Jurassic–Cenozoic tholeiite and calc-alkaline rocks (e.g., Smellie et al., 1984). Much of King George Island consists of Late Jurassic and Palaeogene igneous rocks (Barton, 1965). The oldest of these are thick successions of andesite lavas that have been correlated with Late Jurassic volcanic accumulations on the Antarctic Peninsula. The Fildes Peninsula Group, which (as the name implies) is largely confined to the Fildes Peninsula, includes layers of andesite, agglomerate and tuff with

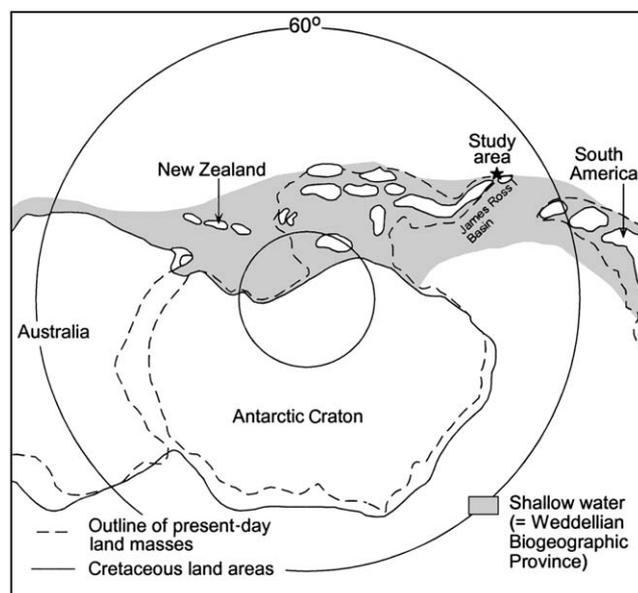


Fig. 8. Weddellian Biogeographic Province during the Late Cretaceous and Early Cenozoic (based on Case, 1988, fig. 6).

interbedded sedimentary rocks of Palaeogene age (Watts, 1982; Weaver et al., 1982). Late Cretaceous palynofloras have, however, been reported from the island (Dutra and Batten, 2000), and the limited deposits of this age may have contributed in a small way to the recycled assemblages of the AB2 core.

Deposition of the sediments that comprise the S7 and S4 cores took place on the continental slope mainly by mass flow and ice-rafting, and less often by water currents (Powell, 1984). Core S7 is divisible into two parts: an upper section rich in palynomorphs and a lower zone of impoverished assemblages; both contain Late Cretaceous and Palaeogene taxa (Fig. 2). The change from poor to rich assemblages suggests a change from shallow to deeper depositional conditions, the former reflecting a sea-level lowstand resulting from a cold period of glacial advance. This is supported by the presence of gravels in, and the absence of dinoflagellate cysts (Byun et al., 1996) from, the lower section.

The relatively diverse palynomorph assemblages and the abundance of recycled Late Cretaceous and Palaeogene palynomorphs in Core S4 (Fig. 3) could be taken to imply derivation from several areas and transport to the site of deposition by a variety of means. However, most of the sediment is considered to have been deposited by mass flows during a relatively short period because the core is from a steep (5–10°) marine slope to the south of the South Shetlands Islands (Prieto et al., 1997; Yoon, S. et al., 2002) (Fig. 1A).

On the western side of Admiralty Bay, King George Island, from which Core AB2 was obtained, the weathered Paleocene–Eocene basalts of the Fildes Peninsula Group crop out. On its eastern side Eocene–Oligocene fine-grained or glassy hypersthene-augite andesites of the Henequene Formation occur (Weaver et al., 1982). Overall, the taxonomically rich recycled palynomorph assemblages recorded from the core (Fig. 4) are closely comparable to the Campanian–Palaeogene assemblages that have been reported from James Ross (Askin, 1983) and Seymour islands (Askin, 1988a,b, 1989, 1990a,b), and from the exposures of Late Cretaceous deposits on King George Island itself (Dutra and Batten, 2000).

The Late Cretaceous (Campanian–Maastrichtian) dinoflagellate cyst assemblages, which include *Amphidiadema denticulata*, *Isabelidinium thomasii*, *Manumiella? cretacea*, *Odontochitina spinosa*, *Palaeoperidinium pyrophorum* and *Satyrodinium haumuriense* (Byun et al., 1996; nomenclature here according to Fensome and Williams, 2004; see Table 2), are comparable to those of the Santa Marta Formation on James Ross and Humps islands (Sumner, 1992; Wood and Askin, 1992), and the Palaeogene assemblages of *Areosphaeridium diktyoplokum*, *Cordosphaeridium fibrospinosum*, *Enneadocysta multicornuta*, *Isabelidinium pellucidum*, *Senegalinium? dilwynense*, *Vozzhennikovia apertura* and other taxa are similar to those of the Paleocene Sobral Formation on Seymour Island and Eocene La Meseta Formation on Snow Hill Island (Askin, 1988a).

Overall, the composition of both the marine and non-marine palynological assemblages suggests that the sediments

Table 2

Dinoflagellate cysts and an acritarch (asterisked) recorded from the core samples examined, with author attributions and year of publication; nomenclature according to Fensome and Williams (2004) in which details of the publications concerned are provided

<i>Amphidiadema denticulata</i> Cookson and Eisenack, 1960a
<i>Areosphaeridium diktyoplokum</i> (Klumpp, 1953) Eaton, 1971
<i>Beringiella fritilla</i> Bujak, 1984*
<i>Cassidinium fragile</i> (Harris, 1965) Drugg, 1967
<i>Chatangiella</i> sp.
<i>Cordosphaeridium fibrospinosum</i> Davey and Williams, 1966b, emend. Davey, 1969c
<i>Diconodinium cristatum</i> Cookson and Eisenack, 1974, emend. Morgan, 1977
<i>Enneadocysta multicornuta</i> (Eaton, 1971), Stover and Williams, 1995
<i>Impagidinium pallidum</i> Bujak, 1984
<i>Impagidinium patulum</i> (Wall, 1967) Stover and Evitt, 1978
<i>Isabelidinium</i> sp.
<i>Isabelidinium pellucidum</i> (Deflandre and Cookson, 1955) Lentin and Williams, 1977a
<i>Isabelidinium thomasii</i> (Cookson and Eisenack, 1961a) Lentin and Williams, 1977a
<i>Laciniadinium arcticum</i> (Manum and Cookson, 1964) Lentin and Williams, 1980
<i>Operculodinium centrocarpum</i> (Deflandre and Cookson, 1955) Wall, 1967
<i>Manumiella? cretacea</i> (Cookson, 1956) Bujak and Davies, 1983
<i>Nelsoniella tuberculata</i> Cookson and Eisenack 1960a
<i>Manumiella seelandica</i> (Lange, 1969) Bujak and Davies, 1983, emend. Firth, 1987
<i>Nelsoniella tuberculata</i> Cookson and Eisenack, 1960a
<i>Odontochitina spinosa</i> Wilson, 1984c
<i>Operculodinium centrocarpum</i> (Deflandre and Cookson, 1955) Wall, 1967
<i>Paleoperidinium pyrophorum</i> (Ehrenburg, 1838 ex Wetzel, 1933a) Sarjeant, 1967b
<i>Satyrodinium haumuriense</i> (Wilson, 1984c) Lentin and Manum, 1986
<i>Senegalinium</i> sp.
<i>Senegalinium? dilwynense</i> (Cookson and Eisenack, 1965c) Stover and Evitt, 1978
<i>Vozzhennikovia apertura</i> (Wilson, 1967a) Lentin and Williams, 1976

examined came from the Weddell Sea rather than from the Bellingshausen Sea, mostly transported via ice-rafted debris or by discharge of subglacial meltwater (Fig. 9). The sedimentary formations that crop out on the eastern side of the Antarctic Peninsula, including those on James Ross, Seymour, Joinville and Snow Hill Islands, are considered to have been the main sources of the recycled material.

5.3. Evolution and migration of angiosperm pollen

It is clear from the palynological and plant megafossil record that during the Late Cretaceous and Palaeogene the vegetation of the Antarctic Peninsula was much richer and more varied than today, and that it became increasingly impoverished as the climate deteriorated from the Eocene onwards (Elliot, 1985; Truswell, 1991; Hansom and Gordon, 1998), although even during the mid Late Pliocene woody angiosperms still inhabited inland Antarctica, suggesting the overall presence of a complex and diverse vegetation (Hill, 1991) that was finally obliterated during the onset of the Pleistocene glaciation.

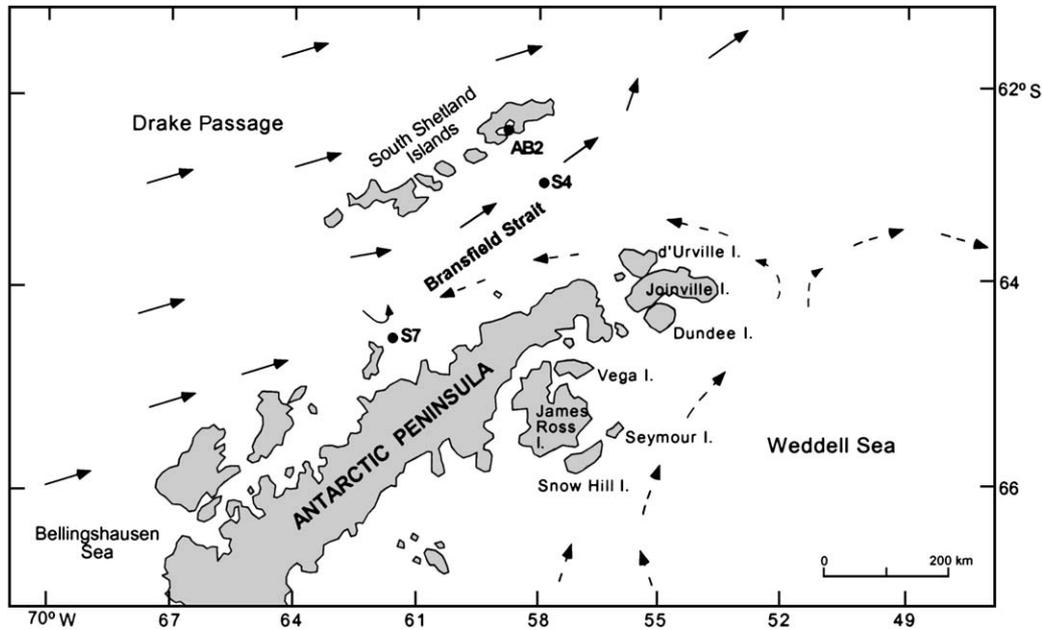


Fig. 9. Main surface currents in Bransfield Strait and vicinity. Dashed arrows indicate cold, saline Weddell Sea water; solid arrows indicate relatively warm and less saline water masses originating from the Bellingshausen Sea (cf. Byun et al., 1996, fig. 5, Bahk et al., 2003, fig. 1, and others). Arrows to the north of the South Shetland Islands indicate the Antarctic Circumpolar Current.

Angiosperms migrated into Antarctica from South America during the Cretaceous Period, and possibly also from south-east Asia via Australia (Dettmann and Jarzen, 1988; Askin, 1989; Truswell, 1990; Hill and Scriven, 1995). They speciated rapidly at these high latitudes, contributing to the development of the angiosperm vegetation that dominates the Southern Hemisphere today. The migration and evolution of early angiosperms in Gondwana was probably enhanced by the high level of disturbance caused primarily by the rifting of the supercontinent (Truswell, 1990; Hill and Scriven, 1995). The recycled palynofloras of the Bransfield Strait appear to be typical of the Weddellian Biogeographic Province. As the climate deteriorated during the Cenozoic, the angiosperm flora was reduced in biomass and diversity, finally being restricted to the remnants that are present today (Francis, 1991; Hill and Scriven, 1995).

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